

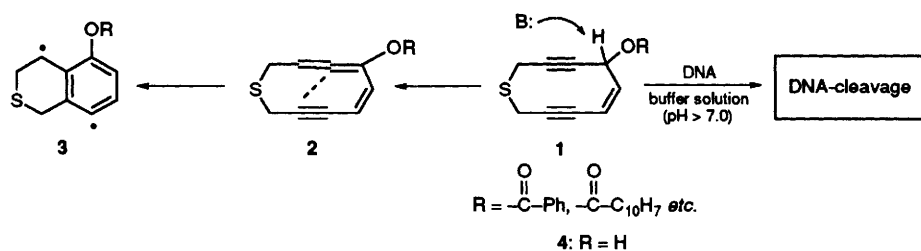
## Design, Synthesis and DNA Cleaving Profiles of Hybrids Containing the Novel Eneidyne and Naturally Occurring DNA Intercalators

Kazunobu Toshima,\* Kazumi Ohta, Aya Ohashi, Takatsugu Nakamura, Masaya Nakata and Shuichi Matsumura  
 Department of Applied Chemistry, Keio University, 3-14-1 Hiyoshi, Kohoku-ku, Yokohama 223, Japan

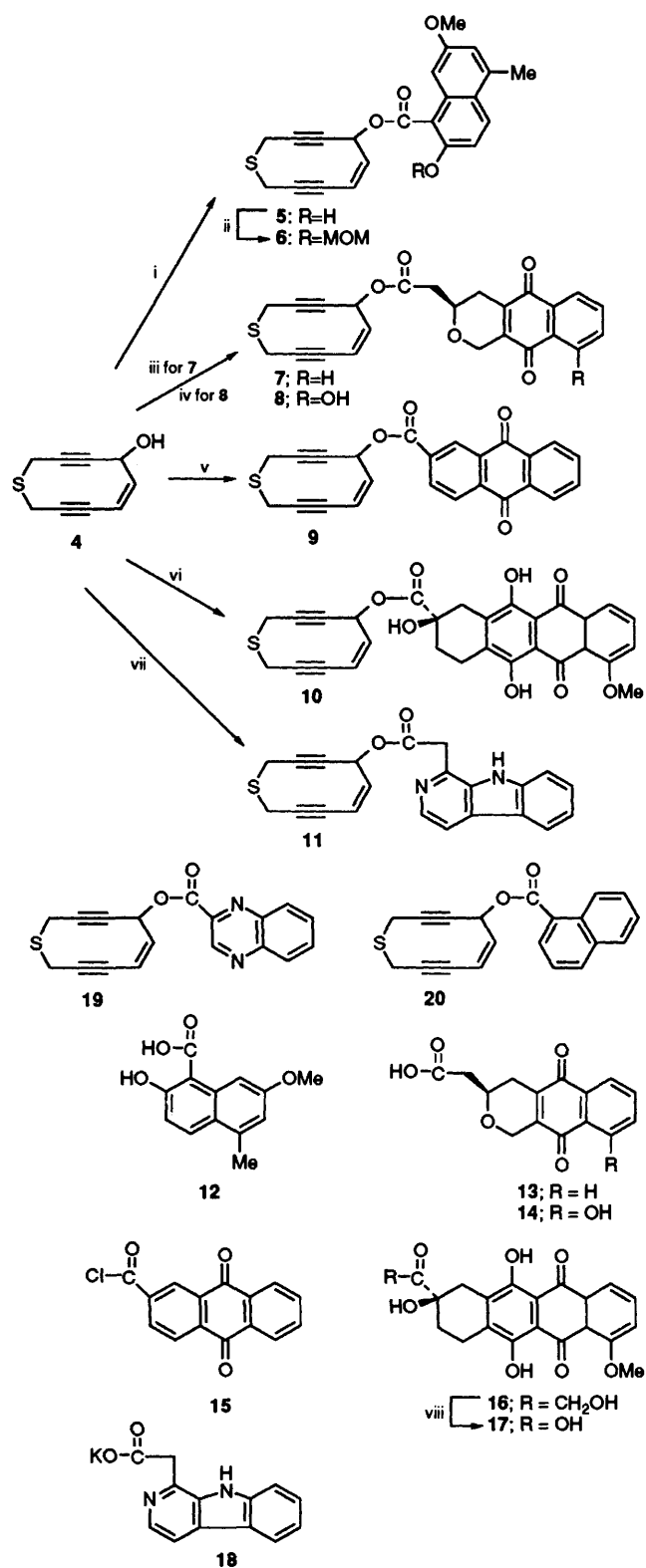
The designed hybrids **5–11** containing the DNA cleavable enediynes **4** and naturally occurring DNA intercalators **12–18** cleave DNA effectively under alkaline conditions without any additive; the hybrids **5** and **11** exhibit the strongest DNA-cleaving abilities with the identical high purine base (G>A) selectivities for their DNA-cleavage profiles.

After the discovery of enediyne antitumour antibiotics, much effort has been paid to developing simple and stable molecules, which generate benzenoid diradical species under specific conditions to cleave DNA, in bioorganic chemistry,

molecular biology and pharmacology, as well as in organic synthesis.<sup>1</sup> Earlier studies in our laboratories have shown that the simple 10-membered heterocyclic enediyne **1** is quite stable when handled at ambient temperature and produced



Scheme 1



**Scheme 2** Reagents and conditions: i, **12** (1.0 equiv.), EDAC (1.0 equiv.), CH<sub>2</sub>Cl<sub>2</sub> 26 °C, 1 h, 45%; ii, MeOCH<sub>2</sub>Cl (3 equiv.), Pr<sub>2</sub>EtN (3.5 equiv.), CH<sub>2</sub>Cl<sub>2</sub>, 26 °C, 24 h, 82%; iii, **13** (1.0 equiv.), 2,4,6-trichlorobenzoyl (TCB<sub>3</sub>Cl) chloride (1.0 equiv.), Et<sub>3</sub>N (1.0 equiv.), 4-dimethylaminopyridine (DMAP) (4.0 equiv.), CH<sub>2</sub>Cl<sub>2</sub>, 26 °C, 2 h, 50%; iv, **14**, (1.0 equiv.), TCB<sub>3</sub>Cl (1.0 equiv.), Et<sub>3</sub>N (1.0 equiv.), DMAP (4.0 equiv.), CH<sub>2</sub>Cl<sub>2</sub>, 26 °C, 2 h, 45%; v, **15** (1.2 equiv.), Et<sub>3</sub>N (1.5 equiv.), CH<sub>2</sub>Cl<sub>2</sub>, 26 °C, 1 h, 90%; vi, **17** (1.0 equiv.), Ph<sub>3</sub>P (3 equiv.), diethyl azodicarboxylate (3.0 equiv.), tetrahydrofuran (THF), 26 °C, 15 min, 17%; vii, **18** (1 equiv.), TCB<sub>3</sub>Cl (1.0 equiv.), Et<sub>3</sub>N (2.0 equiv.), DMAP (0.3 equiv.), THF, 26 °C, 1 h, 19%; viii, HIO<sub>4</sub> (2.0 equiv.), dioxane-H<sub>2</sub>O, 26 °C, 2 h, 87%

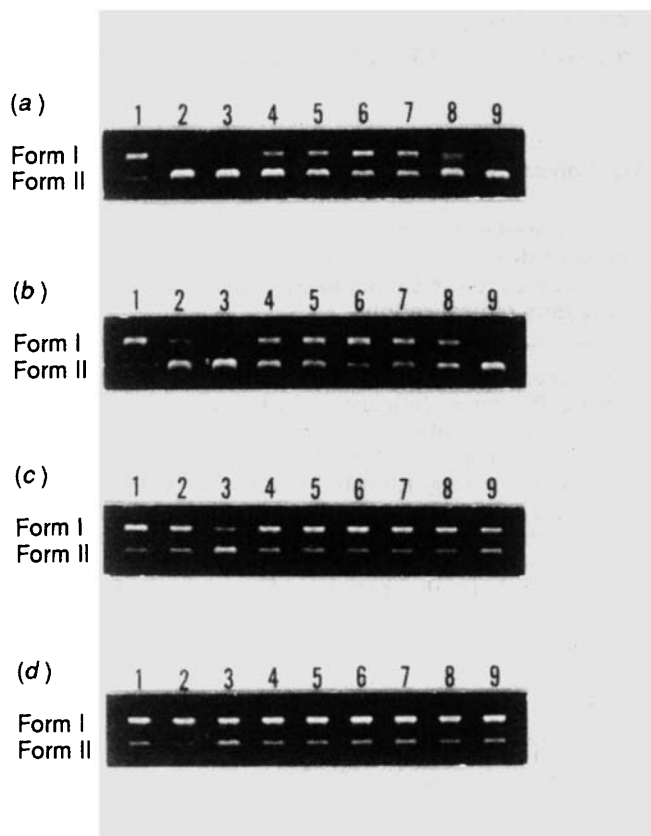
the benzenoid diradical **3** through the enyne-allene **2** under basic conditions.<sup>2</sup> From the preliminary experiments, it was found that **4** (R = H) had a slight DNA-cleaving activity under basic conditions without any additive but its activity could be improved by the introduction of a simple DNA intercalative moiety<sup>3</sup> (Scheme 1). Therefore, hybridization to **4** with a naturally occurring DNA intercalator would be expected to produce both the higher DNA-cleaving ability and base- and sequence-selectivities. Here, we report the syntheses of the hybrid molecules **5–11** containing the novel enediyne system **4** and the DNA intercalator occurring in several bioactive natural products<sup>4</sup> and their DNA cleaving profiles.

As the DNA intercalator, we selected several aromatic moieties occurring in neocarzinostatin<sup>5</sup> (enediyne antibiotic), nanaomycin<sup>6</sup> (pyranonaphthoquinone antibiotic), adriamycin<sup>7</sup> (anthracycline antibiotic) or manzamine<sup>8</sup> (alkaloid). The syntheses of these hybrids are summarized in Scheme 2. The hybrid **5** containing the naphthoate moiety of neocarzinostatin was synthesized from **4** and 2-hydroxy-7-methoxy-5-methylnaphthalene-1-carboxylic acid **12**.<sup>9</sup> The enediyne **4** was effectively esterified with **12** in the presence of 1-ethyl-3-(dimethylaminopropyl)carbodiimide (EDAC) hydrochloride in CH<sub>2</sub>Cl<sub>2</sub> to give **5**. The methoxymethyl (MOM)-protected analogue **6** was prepared from **5** and MeOCH<sub>2</sub>Cl to examine the effect of the hydroxy group of **5** on the DNA-cleaving ability. The pyranonaphthoquinone derivatives **7** and **8** were obtained by esterification of **4** with **13**<sup>†</sup> and **14**<sup>†</sup>, respectively, using the Yamaguchi method.<sup>10</sup> Both carboxylic acids were prepared according to Tatsuta's nanaomycin synthesis procedures.<sup>11</sup> The anthraquinone **9** was obtained by esterification of **4** with 2-anthraquinonecarbonyl chloride **15** using Et<sub>3</sub>N as the base. The hybrid **10** possessing the anthracycline moiety of adriamycin was also synthesized by esterification of **4** with **17** via the Mitsunobu reaction.<sup>12</sup> The carboxylic acid **17** was readily obtained from natural adriamycin through oxidative cleavage of the α-hydroxy ketone **16**<sup>13</sup> by HIO<sub>4</sub> in dioxane-H<sub>2</sub>O. The hybrid **11** having a β-carboline moiety of the antitumour alkaloid, manzamine, was synthesized by esterification of **4** with the β-carboline carboxylic acid potassium salt **18**<sup>14</sup> using the Yamaguchi method.

The DNA-cleaving activities of these compounds and compound **19**<sup>3</sup> as a control are shown in Fig. 1. Although the enediyne **4** itself had a very weak DNA-cleaving activity at 1000 μmol dm<sup>-3</sup>,<sup>3</sup> all the hybrids **5–11** clearly cleaved the covalently closed supercoiled ΦX174 DNA (form I) to the open circular DNA (form II) at pH 8.5 and at 37 °C without any additive in concentrations from 100 to 1000 μmol dm<sup>-3</sup> [(a) and (b) in Fig. 1]. Remarkably, the hybrids **5** and **11** exhibited the strongest DNA-cleaving abilities and caused DNA breaks even at 1–10 μmol dm<sup>-3</sup> [(c) and (d) in Fig. 1]. The potency of **5** and **11** was outstanding among the reported enediyne-based nonnatural systems.<sup>1</sup> This result suggested that the β-carboline part of manzamine was a good DNA intercalator like the naphthoate moiety of neocarzinostatin. Furthermore, it was confirmed that the hydroxy group of the naphthoate moiety of neocarzinostatin played an important role in DNA-cleaving ability (cf. lane 3 with lane 4 in Fig. 1). Their DNA-cleavage site specificity was also analysed. Fig. 2 shows the DNA cleavage results with hybrids **5**, **7**, **10**, **11**, **19** and **20**<sup>3</sup> and the singly 5'-end <sup>32</sup>P labelled double-stranded M13mp18 DNA. Comparisons of the cleavage products with both the enzymatically produced Sanger markers<sup>15</sup> and the

<sup>†</sup> The synthetic procedure of the compound will be reported elsewhere in detail.

<sup>‡</sup> Since the Sanger sequencing reactions result in base incorporation, cleavage at nucleotide N (sequencing) represents cleaving site by a hybrid of Maxam-Gilbert reaction at N + 1. D. L. Boger, S. A. Munk, H. Zarrinmayeh, T. Ishizaki, J. Haught and M. Bina, *Tetrahedron*, 1991, **47**, 2661.



**Fig. 1** Supercoiled DNA cleavage by the hybrids 5–11.  $\Phi$ X174 form I DNA ( $50 \text{ mmol dm}^{-3}$  base pair) was incubated for 24 h with various compounds (a)  $1000 \text{ } \mu\text{mol dm}^{-3}$  at  $37^\circ\text{C}$ , (b)  $100 \text{ } \mu\text{mol dm}^{-3}$  at  $37^\circ\text{C}$ , (c)  $10 \text{ } \mu\text{mol dm}^{-3}$  at  $37^\circ\text{C}$  and (d)  $1 \text{ } \mu\text{mol dm}^{-3}$  at  $42^\circ\text{C}$  in 20% dimethyl sulfoxide in tris-acetate buffer (pH 8.5,  $50 \text{ mol dm}^{-3}$ ) and analysed by electrophoresis (1% agarose gel, ethidium bromide stain). Lane 1: DNA alone; lanes 2–9 correspond to compounds 19, 5–11, respectively.

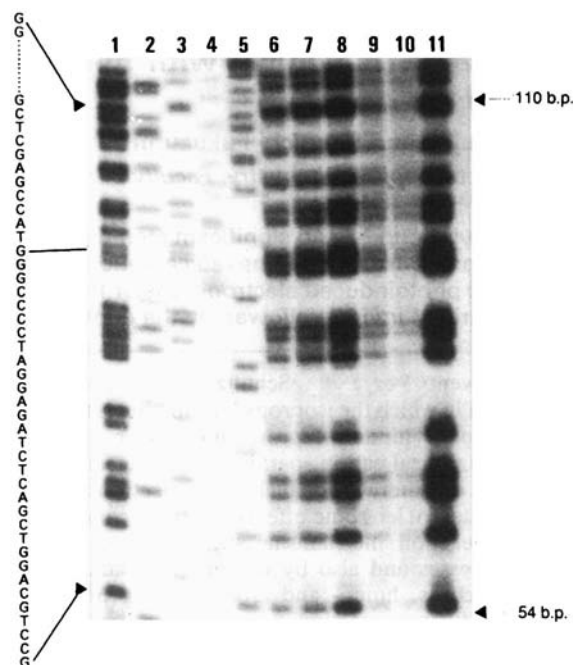
chemically produced Maxam–Gilbert (A + G) marker<sup>16</sup> clearly indicated the identical high purine base (G>A) selectivity of these compounds for their DNA-cleavage profiles. Unexpectedly, the base selectivity was highly independent of the DNA intercalator examined even in the case of the hybrid 5, which had the DNA recognition moiety (T>A>>C>G) of the neocarzinostatin chromophore,<sup>17</sup> and its selectivity was similar to that of the A+G-selective alkylating agents. These results strongly suggested that these intercalators in 5, 7, 10, 11, 19 and 20 significantly increased the DNA cleaving activity, but neither the base nor sequence selectivity. The base selectivity of the hybrid molecules seems to only reflect the reactivity of the enediyne moiety 4 towards purine bases (G>A) like a alkylation mechanism.<sup>18</sup> Details of the DNA cleavage mechanism and the base specificity of these hybrid molecules are now under investigation.

We are grateful to the Institute of Microbial Chemistry for the generous support of our program. Financial supports by The Kurata Foundation and Terumo Life Science Foundation are gratefully acknowledged.

Received, 14th June 1993; Com. 3/034111

## References

- 1 K. C. Nicolaou and W.-M. Dai, *Angew. Chem., Int. Ed. Engl.*, 1991, **30**, 1387.
- 2 K. Toshima, K. Ohta, T. Ohtake and K. Tatsuta, *Tetrahedron Lett.*, 1991, **32**, 391.
- 3 K. Toshima, K. Ohta, A. Ohashi, A. Ohtsuka, M. Nakata and K. Tatsuta, *J. Chem. Soc., Chem. Commun.*, 1992, 1306.
- 4 For synthesis of a hybrid of enediyne and DNA binder: M. Hiram, T. Gomibuchi, K. Fujiwara, Y. Sugiura and M. Uesugi, *J. Am. Chem. Soc.*, 1991, **113**, 9851; K. C. Nicolaou, E. P. Schreiner and W. Stahl, *Angew. Chem., Int. Ed. Engl.*, 1991, **30**, 585; M. Tokuda, K. Fujiwara, T. Gomibuchi, M. Hiram, M. Uesugi and Y. Sugiura, *Tetrahedron Lett.*, 1993, **34**, 669.
- 5 N. Ishida, K. Miyazaki, K. Kumagai and M. Rikimaru, *J. Antibiot.*, 1965, **18**, 68; K. Edo, M. Mizugaki, Y. Koide, H. Seto, K. Furihara, N. Otake and N. Ishida, *Tetrahedron Lett.*, 1985, **26**, 331.
- 6 S. Omura, H. Tanaka, Y. Okada and H. Marumo, *J. Chem. Soc., Chem. Commun.*, 1976, 320; S. Omura, H. Tanaka, Y. Koyama, R. Oiwa, M. Katagiri, J. Awaya, T. Nagai and T. Hata, *J. Antibiot.*, 1974, **27**, 363; H. Tanaka, Y. Koyama, T. Nagai, H. Marumo and S. Omura, *J. Antibiot.*, 1975, **28**, 868.
- 7 F. Arcamone, G. Franceschi, S. Penco and A. Selva, *Tetrahedron Lett.*, 1969, 1007; R. H. Blum and S. K. Carter, *Ann. Intern. Med.*, 1974, **80**, 249; S. K. Carter, *J. Natl. Cancer Inst.*, 1975, **55**, 1265.
- 8 R. Sakai and T. Higa, *J. Am. Chem. Soc.*, 1986, **108**, 6404; R. Sakai, S. Kohmoto and T. Higa, *Tetrahedron Lett.*, 1987, **28**, 5493.
- 9 M. Shibuya, K. Toyooka and S. Kubota, *Tetrahedron Lett.*, 1984, **25**, 1171; K. Shishido, A. Yamashita, K. Horoya, K. Fukumoto and T. Kametani, *Tetrahedron Lett.*, 1989, **30**, 111; K. Takahashi, T. Suzuki and M. Hiram, *Tetrahedron Lett.*, 1992, **33**, 4603.
- 10 J. Inagawa, K. Hirata, H. Saeki, T. Katsuki and M. Yamaguchi, *Bull. Chem. Soc. Jpn.*, 1979, **52**, 1989.
- 11 K. Tatsuta, K. Akimoo, M. Annaka, Y. Ohno and M. Kinoshita, *Bull. Chem. Soc. Jpn.*, 1985, **58**, 1699.
- 12 T. Kurihara, Y. Nakajima and O. Mitsunobu, *Tetrahedron Lett.*, 1976, 2455.
- 13 T. H. Smith, A. N. Fujiwara, W. W. Lee, H. Y. Wu and D. W. Henry, *J. Org. Chem.*, 1977, **42**, 3653.
- 14 Y. Torisawa, A. Hashimoto, M. Nakagawa, H. Seki, R. Hara and T. Hino, *Tetrahedron Lett.*, 1991, **47**, 8067.
- 15 F. Sanger, S. Nicklen and A. R. Coulson, *Proc. Natl. Acad. Sci. USA*, 1977, **74**, 5463.
- 16 A. M. Maxam and W. Gilbert, *Methods Enzymol.*, 1980, **65**, 449.
- 17 A. Galat and I. H. Goldberg, *Nucleic Acids Res.*, 1990, **18**, 2093.
- 18 K. C. Nicolaou, S. Wendeborn, P. Malignes, K. Isshiki, N. Zein and G. Ellestad, *Angew. Chem., Int. Ed. Engl.*, 1991, **30**, 418.



**Fig. 2** Autoradiogram of 12% polyacrylamide-8 mol  $\text{dm}^{-3}$  urea slab gel electrophoresis for sequence analysis. The 5'-end-labelled M13mp18 DNA was cleaved by hybrids 5, 7, 10, 11, 19, and 20 at pH 8.5 and at  $45^\circ\text{C}$  for 24 h (bases 54–110 are shown). Lane 1: Maxam–Gilbert AG reaction; lanes 2–5: Sanger A, G, C and T reactions, respectively; lanes 6–11: 20 (2), 19 (2), 5 (1), 7 (1), 10, (2) and 11 (2 mmol  $\text{dm}^{-3}$ ), respectively.